

# Role of Amino Acids in Nitrogen Fixation during the *R. leguminosarum*/Pea Symbiosis

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## INTRODUCTION

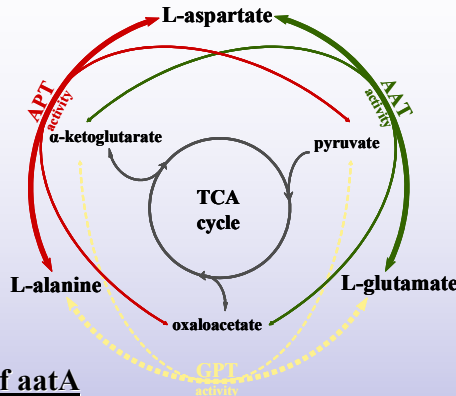
The possible cycling of amino acids between plant and bacteroid has been proposed to be essential for nitrogen assimilation (Lodwig et al., (2003) Nature 422:722-726). Amino acid uptake by *R. leguminosarum* is dominated by two ABC transporters, the general amino acid permease (aap) and the branched amino acid permease (bra). An aap/bra double mutant is  $Fix_{reduced}$  as measured by Nitrogenase activity. However, the nodules are pink compared to the white nodules of a true  $Fix^-$  or red nodules of the wild type. The plants are nitrogen starved as observed by plant dry weight and total nitrogen content analysis.

To explain this we proposed a model, whereby amino acids are required to cycle between the bacteroid and the plant. In this model an amino acid such as glutamate as well as a  $C_4$ -dicarboxylic acid (e.g. L-malate), is taken up by the bacteroid to drive secretion of amino acids such as aspartate or alanine.

## Role of the Aminotransferases

Since aminotransferases link the TCA cycle and amino acid synthesis, we expected they would have a major effect on nitrogen fixation. A mutant in the enzyme aspartate aminotransferase (aatA) formed defective nodules that are not able to fix nitrogen. The role of aatA would be the transamination of oxaloacetate by glutamate to form aspartate (named AAT activity). Furthermore, we have found that *R. leguminosarum*, and possibly all bacteria, lack the classical alanine aminotransferase activity (widely distributed in eukaryotes) which links glutamate and alanine (named GPT activity).

Equally remarkably, we have discovered that aatA catalyses a second transamination reaction in which aspartate will transaminate pyruvate to form alanine (named APT activity). Thus, the glutamate, aspartate and alanine pools in *R. leguminosarum* are inter-connected by the one enzyme (aatA). This has profound consequences for the regulation of bacteroid metabolism and amino acid shuttling.



## Purification and Kinetics of aatA

AatA has been cloned into pET101/D. The plasmid transformed and expressed into BL21 Star™ (DE3). The enzyme is his-tagged, allowing a simple two step purification with a His-Bind® Resin loaded column and a Sephacryl™ S-200.

	AAT activity ( $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}$ $\text{prot.}^{-1}$ )	Total AAT activity ( $\mu\text{mol}\cdot\text{min}^{-1}$ )
Crude Extract	$3.00 \pm 0.34$	$1103 \pm 125$
Pure aatA	$163 \pm 13.4$	$284 \pm 23.5$

Table 2: yield of aatA purification



Fig 5: aatA purification

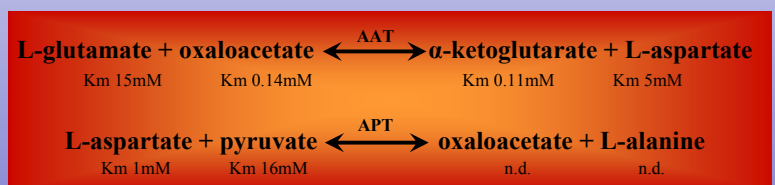


Fig 6: aatA affinities



Fig 1: effect of aap/bra mutant on plants



A34 (wt)



RU1357 (aap bra mutant)

Fig 2: close-up of nodules

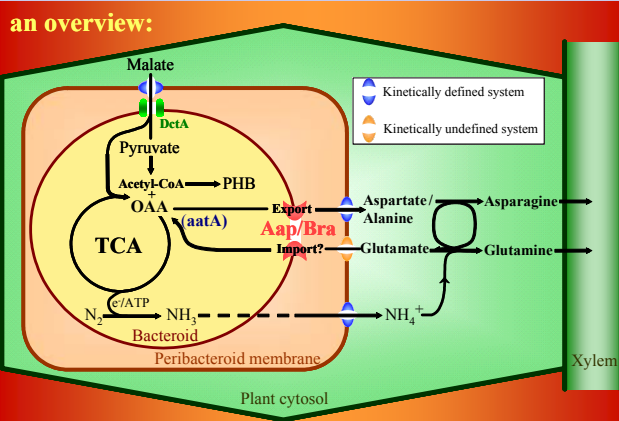


Fig 3: model of plant/bacteroid amino acid cycle



Fig 4: effect of aatA mutant and complemented mutant on plants

Strain	Growth conditions	AAT activity ( $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}$ $\text{prot.}^{-1}$ )	APT activity ( $\mu\text{mol}\cdot\text{min}^{-1}\cdot\text{mg}$ $\text{prot.}^{-1}$ )
A34	Glucose/NH4	$0.299 \pm 0.125$	$0.075 \pm 0.004$
A34	Succinate/NH4	$0.451 \pm 0.157$	$0.194 \pm 0.056$
RU1640 (aatA mutant)	Succinate/NH4	$0.020 \pm 0.005$	$0.013 \pm 0.002$
RU1640 pJP2 aatA (complemented)	Succinate/NH4	$1.600 \pm 0.835$	$0.365 \pm 0.158$
RU1357 (aap/bra mutant)	Succinate/NH4	$0.514 \pm 0.219$	$0.121 \pm 0.076$
Bacteroids	4 week plants	$0.173 \pm 0.015$	$0.049 \pm 0.035$

Table 1: summing up of AAT and APT activities

## Conclusions

- AatA is essential for nitrogen fixation.
- AatA has two activities that allow the conversion of glutamate to aspartate and aspartate to alanine.
- This provides a powerful explanation the origins of the aspartate and of the alanine secreted in various ratios by bacteroids.

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